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Hemochromatosis classification: update and recommendations by the BIOIRON Society

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The recent advances in the knowledge of pathophysiology and molecular basis of iron metabolism have highlighted that HC is caused by mutations in at least five genes, resulting in insufficient hepcidin production or, rarely, resistance to hepcidin action. This has led to an HC classification based on different molecular subtypes, mainly reflecting successive gene discovery. This scheme was difficult to adopt in clinical practice and therefore needs revision. Here we present recommendations for unambiguous HC classification developed by a working group of the International Society for the Study of Iron in Biology and Medicine (BIOIRON Society) including both clinicians and basic scientists during a meeting in Heidelberg, Germany. We propose to deemphasize the use of the molecular subtype criteria in favor of a classification addressing both clinical issues and molecular complexity. Ferroportin Disease (former type 4a) has been excluded because of its distinct phenotype. The novel classification aims to be of practical help whenever a detailed molecular characterization of HC is not readily available.

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Abstract

Hemochromatosis (HC) is a genetically heterogeneous disorder in which uncontrolled intestinal iron absorption may lead to progressive iron overload responsible for disabling and life-threatening complications such as arthritis, diabetes, heart failure, hepatic cirrhosis, and hepatocellular carcinoma.

The recent advances in the knowledge of pathophysiology and molecular basis of iron metabolism have highlighted that HC is caused by mutations in at least five genes, resulting in insufficient hepcidin production or, rarely, resistance to hepcidin action. This has led to an HC classification based on different molecular subtypes, mainly reflecting successive gene discovery. This scheme was difficult to adopt in clinical practice and therefore needs revision. Here we present recommendations for unambiguous HC classification developed by a working group of the International Society for the Study of Iron in Biology and Medicine (BIOIRON Society) including both clinicians and basic scientists during a meeting in Heidelberg, Germany. We propose to deemphasize the use of the molecular subtype criteria in favor of a classification addressing both clinical issues and molecular complexity. Ferroportin Disease (former type 4a) has been excluded because of its distinct phenotype. The novel classification aims to be of practical help whenever a detailed molecular characterization of HC is not readily available.

Historical Perspective

It is commonly accepted that the term "hemochromatosis" was coined by the German pathologist von Recklinghausen in 1889. Of note, this was during the *Versammlung Deutscher Naturforscher* (meeting of German scientists) held in Heidelberg, like the BIOIRON Society (formerly IBIS) meeting in 2019, which led to the current report. Following the description of patients with "bronze diabetes and cirrhosis" by French physicians led by Armand Trousseau in the mid-1800s, von Recklinghausen hypothesized that something circulating in the blood ("Hemo-") was responsible for skin and organ damage and pigmentation ("-chromatosis") (HC). Recognizing excess iron as the etiology of organ toxicity took several decades, and was attributable to Joseph Sheldon in 1935, who was also the first to suggest the genetic origin of the metabolic defect (for a comprehensive historical review, see[1]). Overall, these pioneers' works clearly defined a clinical-pathological entity caused by progressive iron accumulation and characterized by multi-organ damage (mainly in liver, pancreas, joints, heart, and endocrine glands), without signs of anemia (to the contrary, some patients show mildly increased Hb levels)[2-5]. In the 1950s, ferrokinetic studies revealed abnormally increased intestinal iron absorption as the key pathophysiological feature of HC,[6] and repeated/frequent phlebotomies were established as the mainstay of treatment[7]. In 1977, the seminal work by Marcel Simon and colleagues reported the tight linkage between the major histocompatibility complex (MHC) and the putative hemochromatosis gene on chromosome 6p, definitively demonstrating the genetic origin of the disease[8]. This paved the way to the discovery, in 1996, of the "hemochromatosis gene" *HFE* (alias "high Fe", official full name "homeostatic iron regulator")[9], which provided additional information about HC. Initially, it appeared that up to 95% of HC cases could be attributed to homozygosity for a single nucleotide change (845 G→A) causing the substitution of cysteine by tyrosine at amino acid 282 (*p.Cys282Tyr* or C282Y variant)[10-13]. A second *HFE* polymorphism, *p.His63Asp*, was detected, whose minor role became clearer later[14]. The meaning of the *p.Cys282Tyr*/*p.His63Asp* compound heterozygosity is discussed in detail below. The high frequency of *p.Cys282Tyr* homozygosity in the original studies resulted from the inclusion of patients mostly of Northern European ancestry, a region where the variant had originated around 4,000 BC.[15,16] Indeed, the *p.Cys282Tyr* variant, frequent in certain geographical regions,[17] is rare or even absent in large areas of the world, including Asian and African countries, as well as in native Americans.[18,19] Subsequent studies in Southern Europe, in the Mediterranean

area, and in Brazil found that at least one-third of subjects with a defined HC phenotype were negative for *p.Cys282Tyr* at *HFE* genetic testing.[20,21]

Over time, it became evident that the genetic basis of HC was more heterogeneous than initially assumed, and several variants in other iron-controlling genes (collectively referred to as "non-*HFE* genes") were progressively associated with the disorder. These include variants on genes coding for a second receptor for transferrin (*TFR2*),[22-24] ferroportin (*SLC40A1*),[25] hepcidin (*HAMP*),[26,27] and hemojuvelin (*HJV*).[28,29] In particular, the discovery of variants in the *HAMP* and *HJV* genes made it possible to define a severe early-onset (*juvenile*) form of HC, with early cardiac and endocrine impairments, as a molecularly distinct entity.

Hepcidin is the master regulator of iron homeostasis[30-33] and its identification has represented a considerable advance in the comprehension of the pathophysiological mechanisms underlying HC. For detailed reviews on hepcidin discovery, functions and regulation, readers are referred elsewhere[34-37]. Briefly, hepcidin is a small peptide hormone produced by the liver, that negatively controls circulating iron levels. Through interaction with ferroportin[38-40] (its receptor and the only cellular iron exporter so far identified in humans), hepcidin inhibits the absorption of dietary iron in the duodenum and its release by spleen macrophages involved in recycling iron from senescent erythrocytes. Molecular defects causing hepcidin deficiency result in uncontrolled intestinal iron absorption, with progressive iron accumulation in tissues, ultimately leading to HC.[1] In most cases gene defects cause insufficient production of hepcidin, while rarely ferroportin resistance to hepcidin is observed (see below). As illustrated in **Figure 1**, hepcidin regulation by iron is quite complex and involves numerous proteins[41,42] whose alterations can compromise hormone synthesis or function. For this reason, identifying the molecular causes of HC is far from simple and requires a deep knowledge of its pathogenetic basis, which is still not completely clarified. In our view, the term "hemochromatosis" should be reserved for this unique clinical entity caused by genetic lesions that primarily affect the hepcidin-ferroportin system and not used to describe clinically distinct iron overload conditions with other causes (see discussion below).

The current clinical scenario

Unlike in the past, fully expressed and potentially lethal HC (with liver cirrhosis, diabetes, endocrine dysfunction, and heart failure) is seen rarely in current clinical practice.[43] This can be ascribed to increased awareness of the disease, and, mostly, to the

routine assessment of iron biomarkers, particularly serum ferritin. Unfortunately, this is counterbalanced by an increased diagnostic challenge for non-experts in the iron field, primarily due to the lack of specificity of ferritin.

Ferritin is an essentially intracellular protein that serves to store iron safely. It is also present at very low concentration (µg/L) in serum, likely through secretion by macrophages[44]. Normal values usually range from 30 to 200 or 300 µg/L in females and males, respectively. The function of secreted extracellular ferritin remains largely unknown.[45,46] Several common conditions lead to increased serum ferritin levels, including virtually all inflammatory disorders, hepatic cytolysis (e.g., during acute or chronic liver disease), or the metabolic syndrome.[47,48] This translates into a huge number of consultations, overuse of the "first-level" genetic test looking for the presence of the common variants in the *HFE* gene, and even misdiagnosis due to incorrect interpretation of the results.[49]

The glycoprotein transferrin is the extracellular carrier of iron that is detectable at high concentration in blood (g/L; the third most abundant protein after albumin). Transferrin saturation (TSAT) is calculated as the ratio between serum iron and transferrin (multiplied by the correction factor 1.42) or, less reliably, between serum iron and total iron binding capacity, and expressed as a percentage. TSAT is much less requested in clinical practice but is much more informative about a possible diagnosis of HC. Normal TSAT varies between 20 and 45%. It has been estimated that hyperferritinemia with normal TSAT is associated with increased iron stores in less than 10% of cases.[50] Importantly, TSAT elevation is the hallmark of HC. In HC patients, high TSAT reflects the increased pool of circulating iron due to insufficient hepcidin production and typically precedes the rise of serum ferritin by several years.[51] TSAT tends to remain elevated even in subjects effectively iron-depleted by phlebotomies. Occasional reports of normal TSAT in *p.Cys282Tyr* homozygotes with hyperferritinemia should always prompt the search for additional cofactors that raise ferritin, such as metabolic syndrome or alcohol intake.[52,53]

Liver biopsy was once regarded as the gold standard for the diagnosis of HC, because it can reveal iron deposition in hepatocytes with the typical decreasing gradient from the periportal zone (most exposed to iron coming from the gut) to the central-lobular zone in the hepatic acinus. However, in current practice the demonstration of *p.Cys282Tyr* homozygosity along with elevated serum ferritin and TSAT is considered sufficient to make the diagnosis of HC. Liver biopsy remains useful for prognostic purposes in HC patients with serum ferritin levels repeatedly >1,000 µg/L, allowing the early identification of

advanced fibrosis or even subclinical cirrhosis. These conditions require close surveillance for hepatocellular carcinoma even after iron depletion.[51] Nowadays, liver biopsy is seldom performed due to its invasiveness, costs, and the increasing availability of non-invasive tools. Indeed, magnetic resonance imaging (MRI) techniques have largely replaced it for the determination of liver iron concentration (LIC). This is obtained indirectly by using various MRI protocols, for which there is still no consensus on the best one. The choice of the protocol mainly depends on local expertise, as well as on the available equipment and software (for detailed reviews, see [54-56]). Moreover, hepatic transient elastography (Fibroscan) is a reliable non-invasive method for detecting liver fibrosis in HC patients, limiting the need for liver biopsy to those with indeterminate results.[57]

Currently, HC is typically suspected in subjects with no or minimal symptoms, increased serum ferritin levels without alternative explanation, high TSAT, and evidence of increased liver iron stores by MRI. Making the diagnosis at this preclinical early stage[51] has the undoubted advantage of preventing organ damage by a relatively simple and cost-effective treatment (phlebotomy), as well as of allowing normal life expectancy.[58,59]

The HFE genetic test, available in most laboratories, identifies the commonest inherited defect in Caucasians predisposing to *HFE*-related HC, i.e. *p.Cys282Tyr* homozygosity. Therefore, in Caucasians, an HFE genetic test is indicated when high TSAT is confirmed irrespective of parenchymal IO demonstration. *p.Cys282Tyr* homozygosity has a variable and difficult to predict clinical penetrance,[60-64] with several inherited and acquired modifiers potentially contributing to the final phenotype. For this reason, *HFE*-related HC should not be viewed as a simple monogenic disorder, but rather as the complex result of the interplay of environmental, lifestyle, and still unidentified genetic cofactors.[1]

Regarding the compound *p.Cys282Tyr* and *p.His63Asp* heterozygosity, compelling evidence exists that this genotype *per se* is characterized by minimal or no clinical penetrance [65,66]. Thus, it cannot be considered diagnostic for HC (for a detailed discussion see[14]), but at most as a susceptibility factor that can be associated with mild-to-moderate IO only in case of digenic inheritance[67] (see below) or when other predominant causes of liver disease are present, namely non-alcoholic fatty liver disease (NAFLD), alcohol, or HCV. Of note, the latter two are known to cause acquired hepcidin suppression[41,68,69]. According to existing guidelines, whenever a subject with *p.Cys282Tyr*/*p.His63Asp* compound heterozygosity has evidence of IO, a secondary cause of liver disease should be sought and treated[70,71], with phlebotomies possibly considered as an adjunctive treatment. On the other hand, the negative effects of an automatic HC (mis)diagnosis in

p.Cys282Tyr/*p.His63Asp* compound heterozygotes are commonly seen at referral centers. They include patients' and family members' unnecessary anxiety, incomplete prior investigations (e.g., serum ferritin but neither TSAT nor MRI), overlooking of other causes, and/or unnecessary treatment by phlebotomies.

Clinical elements that should raise a definite suspicion of HC are reported in **Table 1**. In Caucasians with a negative first-level *HFE* test (i.e., *p.Cys282Tyr* homozygosity is not detected) and in non-Caucasians, a second-level genetic test should be considered in order to identify rarer variants in the *HFE* or in other genes known to be linked to hepcidin control. In general, these types of HC are less influenced by cofactors, and characterized by a more severe and homogeneous clinical picture appearing at a younger age.[72] Their molecular diagnosis is often complex, since variants in HC genes other than *HFE* are typically private, i.e. restricted to members of only one or a few families. To this end, modern approaches based on Next-Generation-Sequencing (NGS) have greatly expanded the diagnostic possibilities in rarer HC, while at the same time opening enormous challenges of interpretation of the results. NGS is generally available only at referral centers and requires specific expertise to avoid misdiagnosis, with long wait time for results (see below). Nonetheless, treatment of patients with a defined HC phenotype should not be delayed pending the result of the genetic test. Recently, NGS methods have also made it possible to estimate the global prevalence of HFE and non-HFE HC in different populations,[19] as summarized in **Table 2**.

Figure 2 illustrates a possible algorithm for diagnosis of HC, starting from clinical, biochemical and imaging studies to molecular confirmation.

The nomenclature of genetic disorders

HC nomenclature suffers from a common problem of classifying genetic diseases. In contrast to genes, diseases lack a standardized way to review official names and symbols by formal committees (https://ghr.nlm.nih.gov/primer/mutationsanddisorders/naming). Disease nomenclature is often derived from the name(s) of the physician(s) who first described the condition, one major sign or symptom, or the biochemical/genetic underlying defect. However, the growing comprehension of the pathophysiological or molecular mechanisms that regulate diseases, as well as the identification of new phenotypes, may require revision of the initial name by experts, in order to improve its usefulness in clinical practice.

Proper nomenclature, in fact, is an essential prerequisite for clear and effective communication about a particular condition. Ideally, it should unequivocally evoke disorders sharing the same pathogenesis and treatment, eventually helping clinicians to provide an accurate diagnosis and management.

What hemochromatosis is (and what it is not)

Nomenclature and case definition of HC have long been recognized as a potential source of confusion[73], especially when dealing with the report of genetic tests [14]. Among experts, there was a common feeling that the HC classification needs to be revised in view of the increasing awareness and knowledge of IO disorders. To this end, the BIOIRON Society promoted a two-step process. The starting point was the preparation of a survey that was sent to working group participants, including both expert clinicians and basic scientists actively involved in the iron metabolism field, and consisting of nearly all who discovered the hemochromatosis genes and hepcidin. The survey questions and the summary of responses are available in the Supplementary Materials (S1). This was followed by a critical collegial discussion during a specific session of the most recent Biennial Meeting of the BIOIRON Society in Heidelberg. The recommendations reported here are the result of such discussion, where the panelists eventually agreed on the novel classification. As an integral part of the process, the panelists agreed on a robust definition of HC, based on clinical presentation and widely available tools, as a prerequisite for genetic testing. The main clinical, biochemical and imaging studies for the suspicion of HC are reported in **Table 1**. Rigorously speaking, the term “hemochromatosis” should be reserved for a unique genetic clinical-pathological condition characterized by increased TSAT, IO in the liver (but not in the spleen), prevalent involvement of peri-portal hepatocytes with iron-spared Kupffer cells, and signs and/or symptoms associated with IO. The panelists also emphasized that the term “hemochromatosis” itself implies an IO of genetic origin, which is why they would recommend avoiding the unnecessary use of qualifiers such as "hereditary", "genetic" or "primary". Indeed, genetic defects in the hepcidin/ferroportin regulatory axis (caused by variants in hepcidin regulators, the hepcidin gene itself, or in ferroportin) are responsible for inadequate production or activity of hepcidin, or lack of hepcidin responsiveness of ferroportin. Finally, the panelists agreed that the definition of HC should also include the absence of hematological signs of a primary/predominant red blood cell disorder, such as anemia or reticulocytosis (see **Table 1** for some exceptions to this rule). This is needed to distinguish HC from other iron overload conditions, often

referred to as “iron-loading anemias”,[74,75] which are similarly characterized by increased TSAT and are nearly always genetically determined. In these conditions, the hepcidin suppression is caused by factors released by erythropoietin-stimulated erythroblasts (e.g. erythroferrone or ERFE),[76] as a consequence of ineffective erythropoiesis or compensated chronic hemolysis, and not by variants in genes affecting the hepcidin-ferroportin axis. The prototype of this group is non-transfusion dependent thalassemia (NTDT).[77] IO also occurs in transfusion dependent inherited anemias, but in this case, it is mainly due to transfusions *per se*, and hepcidin levels tend to be increased,[78] especially immediately after the transfusion because of suppression of erythropoiesis.[79] **Figure 3** illustrates the main mechanisms underlying the development of IO in hemochromatosis and iron loading anemias. As mentioned above, the majority of iron loading anemias are inherited (for recent comprehensive reviews, see[36,80-82]), including forms caused by variants in the hemoglobin genes, in genes coding for red blood cell enzymes or membrane structures, as well as congenital sideroblastic[81] or dyserythropoietic[82] anemias. Sometimes, variants in genes directly regulating iron transport and utilization (such as *DMT1*, transferrin, ceruloplasmin, and others) may be implicated as well.[80] Finally, IO due to hepcidin inhibition can also occur independently of transfusions in some forms of myelodysplastic syndromes[83], especially those characterized by ringed sideroblasts and increased ineffective erythropoiesis associated with acquired somatic mutations in *SF3B1*[84]. In any case, all these conditions should never be regarded as hemochromatosis because of the distinct pathogenesis and treatment (e.g., phlebotomies are often not feasible).

The former classifications: strengths and shortcomings

The HC classifications reported by authoritative textbooks and recent reviews and guidelines are based on a schema (**Table 3**), in which numbers and letters reflect the chronology of first descriptions of genotype-phenotype correlations.[51,71,85,86] Four types are included, with type 2 and type 4 further subdivided into subtypes A and B. They have the advantage of being very informative from a molecular point of view, and officially endorsed by OMIM (the Online Mendelian Inheritance of Man database), but also present several caveats and inconsistencies. The main limitations of the current classifications are listed below.

a. Poor applicability in clinical practice (Limitations due to costs and lack of widespread expertise).

Apart from the genetic test looking for the common *HFE* variants, the identification of the molecular defect causing rarer forms of HC is currently offered by few laboratories, heterogeneously distributed and scattered worldwide. This requires that patients should travel, or DNA should be sent to referral centers, with inevitable discomforts, delays and costs. Moreover, although the second-level genetic test (mainly based on NGS approach) has recently improved, with gradually decreasing costs, it demands advanced experience for a rigorous interpretation, which can take several weeks.[14,87-89] Moreover, some cases of HC still remain molecularly undiagnosed even after NGS, suggesting the possibility of unknown gene(s) yet to be discovered.[89-91] For this reason, the cooperation between geneticists, bioinformaticians, and clinicians is necessary to resolve the most difficult cases. EuroBloodNet, the network connecting experts on rare hematological diseases, is making great efforts in this direction (for details, see www.eurobloodnet.eu).

***b. Numerical subtypes do not capture the complex molecular pathogenesis of HC*.**

Recent applications of NGS have highlighted that some patients with a provisional diagnosis of *non-HFE* HC cannot be ascribed to any of the numerical subtypes listed in the previous classification (**Table 3**). Essentially there are two reasons:

1) Some show a “digenic” inheritance, deriving from the combination of pathogenic variants in two different genes involved in iron metabolism (e.g., single *p.Cys282Tyr* + heterozygous variants in *HJV*, *HAMP* or *TFR2*).[90,92-95] Although there are still only few cases reported, digenic inheritance must also be considered in cases whose *HFE* genotype *per se* does not fully explain the clinical picture, for example in patients with p.Cys282Tyr homozygosity and very early/severe IO.

2) Others do not display variants in any of the five classical hemochromatosis genes (i.e., *HFE, HAMP, HJV, TFR2,* and *SLC40A1*). Recently, some small case-series[96-98] have reported moderate late-onset IO in patients carrying variants in the *BMP6* gene, encoding one of the major activators of hepcidin expression in response to iron[99]. The role of such variants is still controversial, as they have been detected mostly in patients with a substantial burden of acquired cofactors.[100] Nonetheless, they broaden the spectrum of genetic defects potentially responsible for HC.

c. Former Type 4A HC

Former Type 4A HC actually represents an IO syndrome characterized by distinctive clinical, biochemical, and pathological features, which do not fit the definition of HC.[101,102] They include normal to low TSAT, iron retention in spleen and hepatic macrophages, and,

sometimes, poor tolerance to standard phlebotomies. The underlying molecular defect is the presence of loss-of-function (LOF) variants in the *SLC40A1* gene, that reduce expression or iron export capability of ferroportin at the cell surface. Therefore, iron is trapped inside iron-recycling macrophages (primarily in the spleen), resulting in a reduction of circulating iron, and a tendency to iron-restricted erythropoiesis. The corresponding clinical features are normal-to-low TSAT and, sometimes, the development of mild anemia after phlebotomies. Both these elements are clearly at variance with the case definition of HC according to **Table 1**. Another peculiarity is represented by autosomal dominant inheritance. Taking into consideration all these aspects, many authors have suggested adopting a specific terminology for this condition, such as Ferroportin disease (FD).[14,102] In spite of very high ferritin levels which may be evident even in young-adult subjects, FD phenotype is generally milder than in *HFE*-related HC, possibly because of the lower toxicity of iron accumulation in macrophages as compared to hepatocytes.[103].[102,103]

On the other hand, very rare gain-of-function (GOF) variants in the *SLC40A1* gene lead to ferroportin resistance to hepcidin, and cause IO conditions phenotypically and biochemically indistinguishable from hepcidin-deficient HC (former Type 4B HC).[104] Variants that interfere with hepcidin binding to ferroportin and also impair ferroportin stability or ability to export iron are also possible, potentially leading to a mixed or intermediate phenotype variably influenced by age or environmental factors.

d. Former Type 2 molecular subtypes are not always Juvenile forms and vice versa

As mentioned before, the term *juvenile* hemochromatosis classically designates an early-onset (within the second or third decades of life), fully-expressed HC phenotype showing similar penetrance in both genders and a tendency to present with cardiac and endocrine dysfunctions.[27,105] This phenotype is generally due to variants in *HJV* and *HAMP* genes causing much more severe iron hyperabsorption than the *HFE* mutations. However, recent studies have highlighted some age-overlap at diagnosis between the various molecular subtypes of HC.[72] Therefore, the term *juvenile* HC can be ambiguous if invariably attributed to variants in the *HAMP* and *HJV* genes, because in some of these patients the disease is diagnosed in adulthood. Similarly, the term can be misleading in HC patients with defects in genes other than *HJV* or *HAMP*, but with early-onset severe phenotypes.

New classification of HC proposed by the working group

As a result of the two-step process described previously, the panelists propose a new classification of HC (shown in **Table 4**), addressing both clinical issues (thereby addressing the needs of general clinicians and subspecialists) and molecular precision. The emphasis on clinical features obviates the current challenges represented by second-level genetic testing for detecting rare variants in the *HFE* and non-*HFE* genes, which could lead to delayed diagnosis and treatment. When criteria listed on **Table 1** are fulfilled, the diagnosis of *HFE*-related HC can be made in the presence of *p.Cys282Tyr* homozygosity. If an appropriately investigated patient has an unequivocal HC phenotype without cofactors but is not a *p.Cys282Tyr* homozygote (and this includes compound *p.Cys282Tyr* and *p.His63Asp* heterozygosity or *p.His63Asp* homozygosity), a provisional diagnosis of "molecularly undefined" HC can be made, and phlebotomies started. In this case, quantification of the total amount of iron removed by phlebotomies will serve as an additional marker of IO. The panelists agree that, whenever possible, an accurate molecular characterization remains important in these patients, especially for cascade screening of asymptomatic siblings, or other first-degree relatives. To this end, patients should be referred (or DNA should be sent) to a specialized center. Indeed, second-level genetic tests have limitations that include costs, time-delay, and poor availability in certain regions, and require a high level of expertise for interpretation. Based on NGS results, some cases could be reclassified into *HFE*-related, digenic or non-*HFE* HC (as shown in **Figure 2**).

Based on all the above considerations, we suggest adopting a new, more workable classification of HC (shown in **Table 4**) capable of capturing the growing genetic complexity of HC highlighted by NGS. In fact, the type-numerical classification does not allow assignation of any subtypes to patients with complex genotypes deriving from variants in two genes (“digenic” HC), nor to those who remain undefined after sequencing of known HC genes.

Finally, the panelists agreed to abandon the current terminology of Type 4A and 4B HC, related to LOF and GOF variants in the ferroportin gene, respectively. While Type B is in all respects a form of HC (and it should be referred to as ferroportin-related HC), Type 4A has the unique characteristics we described previously. Thus, it should be definitively renamed as "Ferroportin Disease" and included in inherited rare disorders of iron metabolism other than HC. It is important to recall that ferroportin mutations are characterized by an autosomal dominant inheritance pattern, with important implications for genetic testing of family members.

In summary, the novel classification proposed here is based on a pathophysiological cornerstone (hepcidin deficiency) and a distinct clinical/biochemical phenotype. It recognizes the difficulties of a complete molecular characterization and has the potential of being easily shareable between practicing physicians and referral centers. Avoiding any ambiguity is essential for clear and effective communication, that will facilitate proper diagnosis and treatment of HC.

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Authorship and conflict-of-interest statements

PBr, DG, GP, MM and IC led the panel and conceived the manuscript. DG and FB co-wrote the manuscript. PA, EB-J, PBi, BB, CC, RE, RF, TG, OL, GM, EN, APie, APip, DP, JR, MS, PS, DS, HZ critically revised and edited the manuscript. TG and EN are co-founder of Intrinsic LifeScience, and consultant for Protagonist, Vifor Pharma, Ionis Pharmaceuticals, and Disc Medicine. JR is consultant for Bond Biosciences and Gilead. The other authors declare no competing financial interests.

References

1. Pietrangelo A. Hereditary hemochromatosis: pathogenesis, diagnosis, and treatment. *Gastroenterology*. 2010;139(2):393-408, 408 e391-392.
2. Barton JC, Bertoli LF, Rothenberg BE. Peripheral blood erythrocyte parameters in hemochromatosis: evidence for increased erythrocyte hemoglobin content. *J Lab Clin Med*. 2000;135(1):96-104.
3. Beutler E, Felitti V, Gelbart T, Waalen J. Haematological effects of the C282Y HFE mutation in homozygous and heterozygous states among subjects of northern and southern European ancestry. *Br J Haematol*. 2003;120(5):887-893.
4. Datz C, Haas T, Rinner H, Sandhofer F, Patsch W, Paulweber B. Heterozygosity for the C282Y mutation in the hemochromatosis gene is associated with increased serum iron, transferrin saturation, and hemoglobin in young women: a protective role against iron deficiency? *Clin Chem*. 1998;44(12):2429-2432.
5. McLaren CE, Barton JC, Gordeuk VR, et al. Determinants and characteristics of mean corpuscular volume and hemoglobin concentration in white HFE C282Y homozygotes in the hemochromatosis and iron overload screening study. *Am J Hematol*. 2007;82(10):898-905.
6. Alper T, Savage DV, Bothwell TH. Radioiron studies in a case of hemochromatosis. *J Lab Clin Med*. 1951;37(5):665-675.
7. Crosby WH. A history of phlebotomy therapy for hemochromatosis. *Am J Med Sci*. 1991;301(1):28-31.
8. Simon M, Bourel M, Genetet B, Fauchet R. Idiopathic hemochromatosis. Demonstration of recessive transmission and early detection by family HLA typing. *N Engl J Med*. 1977;297(19):1017-1021.
9. Feder JN, Gnirke A, Thomas W, et al. A novel MHC class I-like gene is mutated in patients with hereditary haemochromatosis. *Nat Genet*. 1996;13(4):399-408.
10. Jazwinska EC. Current status in identifying the haemochromatosis gene. *J Gastroenterol Hepatol*. 1996;11(11):1041-1043.
11. Jazwinska EC, Cullen LM, Busfield F, et al. Haemochromatosis and HLA-H. *Nat Genet*. 1996;14(3):249-251.
12. Jazwinska EC, Powell LW. Hemochromatosis and "HLA-H": definite! *Hepatology*. 1997;25(2):495-496.
13. Cullen LM, Gao X, Easteal S, Jazwinska EC. The hemochromatosis 845 G-->A and 187 C-->G mutations: prevalence in non-Caucasian populations. *Am J Hum Genet*. 1998;62(6):1403-1407.
14. Porto G, Brissot P, Swinkels DW, et al. EMQN best practice guidelines for the molecular genetic diagnosis of hereditary hemochromatosis (HH). *Eur J Hum Genet*. 2016;24(4):479-495.
15. Distante S, Robson KJ, Graham-Campbell J, Arnaiz-Villena A, Brissot P, Worwood M. The origin and spread of the HFE-C282Y haemochromatosis mutation. *Hum Genet*. 2004;115(4):269-279.
16. Cassidy LM, Martiniano R, Murphy EM, et al. Neolithic and Bronze Age migration to Ireland and establishment of the insular Atlantic genome. *Proc Natl Acad Sci U S A*. 2016;113(2):368-373.
17. McCullough JM, Heath KM, Smith AM. Hemochromatosis: Niche Construction and the Genetic Domino Effect in the European Neolithic. *Hum Biol*. 2015;87(1):39-58.
18. Lok CY, Merryweather-Clarke AT, Viprakasit V, et al. Iron overload in the Asian community. *Blood*. 2009;114(1):20-25.
19. Wallace DF, Subramaniam VN. The global prevalence of HFE and non-HFE hemochromatosis estimated from analysis of next-generation sequencing data. *Genet Med*. 2016;18(6):618-626.
20. Piperno A, Sampietro M, Pietrangelo A, et al. Heterogeneity of hemochromatosis in Italy. *Gastroenterology*. 1998;114(5):996-1002.

21. Santos PC, Cancado RD, Pereira AC, et al. Hereditary hemochromatosis: mutations in genes involved in iron homeostasis in Brazilian patients. *Blood Cells Mol Dis*. 2011;46(4):302-307.
22. Camaschella C, Roetto A, Cali A, et al. The gene TFR2 is mutated in a new type of haemochromatosis mapping to 7q22. *Nat Genet*. 2000;25(1):14-15.
23. Roetto A, Daraio F, Alberti F, et al. Hemochromatosis due to mutations in transferrin receptor 2. *Blood Cells Mol Dis*. 2002;29(3):465-470.
24. Girelli D, Bozzini C, Roetto A, et al. Clinical and pathologic findings in hemochromatosis type 3 due to a novel mutation in transferrin receptor 2 gene. *Gastroenterology*. 2002;122(5):1295-1302.
25. Montosi G, Donovan A, Totaro A, et al. Autosomal-dominant hemochromatosis is associated with a mutation in the ferroportin (SLC11A3) gene. *J Clin Invest*. 2001;108(4):619-623.
26. Roetto A, Papanikolaou G, Politou M, et al. Mutant antimicrobial peptide hepcidin is associated with severe juvenile hemochromatosis. *Nat Genet*. 2003;33(1):21-22.
27. Camaschella C, Roetto A, De Gobbi M. Juvenile hemochromatosis. *Semin Hematol*. 2002;39(4):242-248.
28. Papanikolaou G, Samuels ME, Ludwig EH, et al. Mutations in HFE2 cause iron overload in chromosome 1q-linked juvenile hemochromatosis. *Nat Genet*. 2004;36(1):77-82.
29. Lanzara C, Roetto A, Daraio F, et al. Spectrum of hemojuvelin gene mutations in 1q-linked juvenile hemochromatosis. *Blood*. 2004;103(11):4317-4321.
30. Park CH, Valore EV, Waring AJ, Ganz T. Hepcidin, a urinary antimicrobial peptide synthesized in the liver. *J Biol Chem*. 2001;276(11):7806-7810.
31. Pigeon C, Ilyin G, Courselaud B, et al. A new mouse liver-specific gene, encoding a protein homologous to human antimicrobial peptide hepcidin, is overexpressed during iron overload. *J Biol Chem*. 2001;276(11):7811-7819.
32. Andrews NC. Forging a field: the golden age of iron biology. *Blood*. 2008;112(2):219-230.
33. Ganz T, Nemeth E. Hepcidin and iron homeostasis. *Biochim Biophys Acta*. 2012;1823(9):1434-1443.
34. Hentze MW, Muckenthaler MU, Galy B, Camaschella C. Two to tango: regulation of Mammalian iron metabolism. *Cell*. 2010;142(1):24-38.
35. Ganz T. Hepcidin and iron regulation, 10 years later. *Blood*. 2011;117(17):4425-4433.
36. Camaschella C, Nai A, Silvestri L. Iron metabolism and iron disorders revisited in the hepcidin era. *Haematologica*. 2020.
37. Muckenthaler MU, Rivella S, Hentze MW, Galy B. A Red Carpet for Iron Metabolism. *Cell*. 2017;168(3):344-361.
38. Aschemeyer S, Qiao B, Stefanova D, et al. Structure-function analysis of ferroportin defines the binding site and an alternative mechanism of action of hepcidin. *Blood*. 2018;131(8):899-910.
39. Pan Y, Ren Z, Gao S, et al. Structural basis of ion transport and inhibition in ferroportin. *Nat Commun*. 2020;11(1):5686.
40. Nemeth E, Tuttle MS, Powelson J, et al. Hepcidin Regulates Cellular Iron Efflux by Binding to Ferroportin and Inducing Its Internalization. *Science*. 2004;306(5704):2090-2093.
41. Girelli D, Nemeth E, Swinkels DW. Hepcidin in the diagnosis of iron disorders. *Blood*. 2016;127(23):2809-2813.
42. Wang CY, Babitt JL. Liver iron sensing and body iron homeostasis. *Blood*. 2019;133(1):18-29.
43. Deugnier Y, Morcet J, Laine F, et al. Reduced phenotypic expression in genetic hemochromatosis with time: Role of exposure to non-genetic modifiers. *J Hepatol*. 2019;70(1):118-125.
44. Cohen LA, Gutierrez L, Weiss A, et al. Serum ferritin is derived primarily from macrophages through a nonclassical secretory pathway. *Blood*. 2010;116(9):1574-1584.
45. Arosio P, Elia L, Poli M. Ferritin, cellular iron storage and regulation. *IUBMB Life*. 2017;69(6):414-422.
46. Arosio P, Carmona F, Gozzelino R, Maccarinelli F, Poli M. The importance of eukaryotic ferritins in iron handling and cytoprotection. *Biochem J*. 2015;472(1):1-15.
47. Knovich MA, Storey JA, Coffman LG, Torti SV, Torti FM. Ferritin for the clinician. *Blood Rev*. 2009;23(3):95-104.
48. Sandnes M, Ulvik RJ, Vorland M, Reikvam H. Hyperferritinemia-A Clinical Overview. *J Clin Med*. 2021;10(9).
49. Cullis JO, Fitzsimons EJ, Griffiths WJ, Tsochatzis E, Thomas DW, British Society for H. Investigation and management of a raised serum ferritin. *Br J Haematol*. 2018;181(3):331-340.
50. Adams PC, Barton JC. A diagnostic approach to hyperferritinemia with a non-elevated transferrin saturation. *J Hepatol*. 2011;55(2):453-458.

51. Brissot P, Pietrangelo A, Adams PC, de Graaff B, McLaren CE, Loreal O. Haemochromatosis. *Nat Rev Dis Primers*. 2018;4:18016.
52. Adams PC, Reboussin DM, Press RD, et al. Biological variability of transferrin saturation and unsaturated iron-binding capacity. *Am J Med*. 2007;120(11):999 e991-997.
53. Adams PCaBJC. Iron overload and cirrhosis in referred HFE p.C282Y homozygotes with normal transferrin saturation and elevated serum ferritin. *Can Liver Journal*. 2020;3(2):188-193.
54. Golfeyz S, Lewis S, Weisberg IS. Hemochromatosis: pathophysiology, evaluation, and management of hepatic iron overload with a focus on MRI. *Expert Rev Gastroenterol Hepatol*. 2018;12(8):767-778.
55. Henninger B, Alustiza J, Garbowski M, Gandon Y. Practical guide to quantification of hepatic iron with MRI. *Eur Radiol*. 2020;30(1):383-393.
56. Franca M, Carvalho JG. MR imaging assessment and quantification of liver iron. *Abdom Radiol (NY)*. 2020;45(11):3400-3412.
57. Legros L, Bardou-Jacquet E, Latournerie M, et al. Non-invasive assessment of liver fibrosis in C282Y homozygous HFE hemochromatosis. *Liver Int*. 2015;35(6):1731-1738.
58. Niederau C, Fischer R, Sonnenberg A, Stremmel W, Trampisch HJ, Strohmeyer G. Survival and causes of death in cirrhotic and in noncirrhotic patients with primary hemochromatosis. *N Engl J Med*. 1985;313(20):1256-1262.
59. Prabhu A, Cargill T, Roberts N, Ryan JD. Systematic Review of the Clinical Outcomes of Iron Reduction in Hereditary Hemochromatosis. *Hepatology*. 2020;72(4):1469-1482.
60. Beutler E, Felitti VJ, Koziol JA, Ho NJ, Gelbart T. Penetrance of 845G--> A (C282Y) HFE hereditary haemochromatosis mutation in the USA. *Lancet*. 2002;359(9302):211-218.
61. Waalen J, Nordestgaard BG, Beutler E. The penetrance of hereditary hemochromatosis. *Best Pract Res Clin Haematol*. 2005;18(2):203-220.
62. Adams PC, Reboussin DM, Barton JC, et al. Hemochromatosis and iron-overload screening in a racially diverse population. *N Engl J Med*. 2005;352(17):1769-1778.
63. Pilling LC, Tamosauskaite J, Jones G, et al. Common conditions associated with hereditary haemochromatosis genetic variants: cohort study in UK Biobank. *BMJ*. 2019;364:k5222.
64. Allen KJ, Gurrin LC, Constantine CC, et al. Iron-overload-related disease in HFE hereditary hemochromatosis. *N Engl J Med*. 2008;358(3):221-230.
65. Gurrin LC, Bertalli NA, Dalton GW, et al. HFE C282Y/H63D compound heterozygotes are at low risk of hemochromatosis-related morbidity. *Hepatology*. 2009;50(1):94-101.
66. Walsh A, Dixon JL, Ramm GA, et al. The clinical relevance of compound heterozygosity for the C282Y and H63D substitutions in hemochromatosis. *Clin Gastroenterol Hepatol*. 2006;4(11):1403-1410.
67. Biasiotto G, Roetto A, Daraio F, et al. Identification of new mutations of hepcidin and hemojuvelin in patients with HFE C282Y allele. *Blood Cells Mol Dis*. 2004;33(3):338-343.
68. Dostalikova-Cimburova M, Balusikova K, Kratka K, et al. Role of duodenal iron transporters and hepcidin in patients with alcoholic liver disease. *J Cell Mol Med*. 2014;18(9):1840-1850.
69. Girelli D, Pasino M, Goodnough JB, et al. Reduced serum hepcidin levels in patients with chronic hepatitis C. *J Hepatol*. 2009;51(5):845-852.
70. European Association For The Study Of The L. EASL clinical practice guidelines for HFE hemochromatosis. *J Hepatol*. 2010;53(1):3-22.
71. Kowdley KV, Brown KE, Ahn J, Sundaram V. ACG Clinical Guideline: Hereditary Hemochromatosis. *Am J Gastroenterol*. 2019;114(8):1202-1218.
72. Sandhu K, Flintoff K, Chatfield MD, et al. Phenotypic analysis of hemochromatosis subtypes reveals variations in severity of iron overload and clinical disease. *Blood*. 2018;132(1):101-110.
73. Adams PC. Hemochromatosis case definition: out of focus? *Nat Clin Pract Gastroenterol Hepatol*. 2006;3(4):178-179.
74. Camaschella C, Nai A. Ineffective erythropoiesis and regulation of iron status in iron loading anaemias. *Br J Haematol*. 2016;172(4):512-523.
75. Finch C. Regulators of iron balance in humans. *Blood*. 1994;84(6):1697-1702.
76. Coffey R, Ganz T. Erythroferrone: An Erythroid Regulator of Hepcidin and Iron Metabolism. *Hemasphere*. 2018;2(2):e35.
77. Musallam KM, Cappellini MD, Wood JC, Taher AT. Iron overload in non-transfusion-dependent thalassemia: a clinical perspective. *Blood Rev*. 2012;26 Suppl 1:S16-19.
78. Origa R, Galanello R, Ganz T, et al. Liver iron concentrations and urinary hepcidin in beta-thalassemia. *Haematologica*. 2007;92(5):583-588.

79. Pasricha SR, Frazer DM, Bowden DK, Anderson GJ. Transfusion suppresses erythropoiesis and increases hepcidin in adult patients with beta-thalassemia major: a longitudinal study. *Blood*. 2013;122(1):124-133.
80. Brissot P, Bernard DG, Brissot E, Loréal O, Troadec MB. Rare anemias due to genetic iron metabolism defects. *Mutat Res*. 2018;777:52-63.
81. Ducamp S, Fleming MD. The molecular genetics of sideroblastic anemia. *Blood*. 2019;133(1):59-69.
82. Iolascon A, Andolfo I, Russo R. Congenital dyserythropoietic anemias. *Blood*. 2020;136(11):1274-1283.
83. Santini V, Girelli D, Sanna A, et al. Hepcidin levels and their determinants in different types of myelodysplastic syndromes. *PLoS One*. 2011;6(8):e23109.
84. Bondu S, Alary AS, Lefevre C, et al. A variant erythroferrone disrupts iron homeostasis in SF3B1-mutated myelodysplastic syndrome. *Sci Transl Med*. 2019;11(500).
85. Fleming RE, Ponka P. Iron overload in human disease. *N Engl J Med*. 2012;366(4):348-359.
86. Powell LW, Seckington RC, Deugnier Y. Haemochromatosis. *Lancet*. 2016;388(10045):706-716.
87. Manolio TA, Fowler DM, Starita LM, et al. Bedside Back to Bench: Building Bridges between Basic and Clinical Genomic Research. *Cell*. 2017;169(1):6-12.
88. Fernandez-Marmiesse A, Gouveia S, Couce ML. NGS Technologies as a Turning Point in Rare Disease Research , Diagnosis and Treatment. *Curr Med Chem*. 2018;25(3):404-432.
89. Wright CF, FitzPatrick DR, Firth HV. Paediatric genomics: diagnosing rare disease in children. *Nat Rev Genet*. 2018;19(5):325.
90. Badar S, Busti F, Ferrarini A, et al. Identification of novel mutations in hemochromatosis genes by targeted next generation sequencing in Italian patients with unexplained iron overload. *Am J Hematol*. 2016;91(4):420-425.
91. McDonald CJ, Ostini L, Wallace DF, Lyons A, Crawford DH, Subramaniam VN. Next-generation sequencing: Application of a novel platform to analyze atypical iron disorders. *J Hepatol*. 2015;63(5):1288-1293.
92. Merryweather-Clarke AT, Cadet E, Bomford A, et al. Digenic inheritance of mutations in HAMP and HFE results in different types of haemochromatosis. *Hum Mol Genet*. 2003;12(17):2241-2247.
93. Gan EK, Trinder D, Ayonrinde OT, Olynyk JK. Genetics of hereditary hemochromatosis: a clinical perspective. *Expert Rev Endocrinol Metab*. 2009;4(3):225-239.
94. Rametta R, Dongiovanni P, Baselli GA, et al. Impact of natural neuromedin-B receptor variants on iron metabolism. *Am J Hematol*. 2020;95(2):167-177.
95. Lanktree MB, Sadikovic B, Waye JS, et al. Clinical evaluation of a hemochromatosis next-generation sequencing gene panel. *Eur J Haematol*. 2017;98(3):228-234.
96. Piubelli C, Castagna A, Marchi G, et al. Identification of new BMP6 pro-peptide mutations in patients with iron overload. *Am J Hematol*. 2017;92(6):562-568.
97. Daher R, Kannengiesser C, Houamel D, et al. Heterozygous Mutations in BMP6 Pro-peptide Lead to Inappropriate Hepcidin Synthesis and Moderate Iron Overload in Humans. *Gastroenterology*. 2016;150(3):672-683 e674.
98. Alvarenga AM, da Silva NK, Fonseca PFS, et al. Novel mutations in the bone morphogenetic protein 6 gene in patients with iron overload and non-homozygous genotype for the HFE p.Cys282Tyr mutation. *Blood Cells Mol Dis*. 2020;84:102444.
99. Xiao X, Alfaro-Magallanes VM, Babitt JL. Bone morphogenic proteins in iron homeostasis. *Bone*. 2020;138:115495.
100. McDonald CJ, Rishi G, Secondes ES, et al. Evaluation of a bone morphogenetic protein 6 variant as a cause of iron loading. *Hum Genomics*. 2018;12(1):23.
101. Pietrangelo A. The ferroportin disease. *Blood Cells Mol Dis*. 2004;32(1):131-138.
102. Pietrangelo A. Ferroportin disease: pathogenesis, diagnosis and treatment. *Haematologica*. 2017;102(12):1972-1984.
103. Vlasveld LT, Janssen R, Bardou-Jacquet E, et al. Twenty Years of Ferroportin Disease: A Review or An Update of Published Clinical, Biochemical, Molecular, and Functional Features. *Pharmaceuticals (Basel)*. 2019;12(3).
104. Bardou-Jacquet E, Ben Ali Z, Beaumont-Epinette MP, Loreal O, Jouanolle AM, Brissot P. Non-HFE hemochromatosis: pathophysiological and diagnostic aspects. *Clin Res Hepatol Gastroenterol*. 2014;38(2):143-154.
105. De Gobbi M, Roetto A, Piperno A, et al. Natural history of juvenile haemochromatosis. *Br J Haematol*. 2002;117(4):973-979.

106. Beutler E, Griffin MJ, Gelbart T, West C. A previously undescribed nonsense mutation of the HFE gene. *Clin Genet*. 2002;61(1):40-42.
107. Mendes AI, Ferro A, Martins R, et al. Non-classical hereditary hemochromatosis in Portugal: novel mutations identified in iron metabolism-related genes. *Ann Hematol*. 2009;88(3):229-234.
108. Zhang W, Wang X, Duan W, et al. HFE-Related Hemochromatosis in a Chinese Patient: The First Reported Case. *Front Genet*. 2020;11:77.
109. Hamdi-Roze H, Beaumont-Epinette MP, Ben Ali Z, et al. Rare HFE variants are the most frequent cause of hemochromatosis in non-c282y homozygous patients with hemochromatosis. *Am J Hematol*. 2016;91(12):1202-1205.
110. Le Gac G, Congiu R, Gourlaouen I, Cau M, Ferec C, Melis MA. Homozygous deletion of HFE is the common cause of hemochromatosis in Sardinia. *Haematologica*. 2010;95(4):685-687.

Table 1. Main clinical, biochemical and imaging elements for the suspicion of Hemochromatosis

Leading
+ TSAT >45% (mainstay)
+ S-Ferritin >200 µg/L (females) or >300 µg/L (males)
+ imaging evidence of liver IO (MRI* and/or biopsy**)
+ Iron deposits in hepatocytes (if biopsy is performed)
+ Absence of "predominant" acquired risk factors for hepcidin deficiency (e.g., alcohol abuse or end-stage liver disease) and iatrogenic iron overload (e.g. regular transfusions)
+ Absence of hematological signs of a primary red blood cell disorder, such as anemia*** (i.e. Hb>120 g/L in females, >130 g/L in males) and/or reticulocytosis
Not always present
± Signs and/or symptoms associated with IO: - skin pigmentation, asthenia - persistent increase of aminotransferases, hepatomegaly, cirrhosis, hepatocellular carcinoma - joint pain, arthritis, chondrocalcinosis, reduced bone mineral density - diabetes mellitus, hypopituitarism, hypoparathyroidism, hypogonadotropic hypogonadism - cardiomyopathy, heart failure, cardiac arrhythmias

TSAT = transferrin saturation; IO = iron overload; MRI = Magnetic Resonance Imaging.

*Liver Iron Content (LIC) quantification by MRI can be obtained using different protocols, which vary depending on local expertise and equipment. With these limitations, any LIC value higher than the upper normal limit (generally set at 36-40 µmol/g dry weight) should lead to consideration of phlebotomies in HC patients. Similarly, LIC >100-120 µmol/g and >240-300 µmol/g are generally considered as overt and severe IO, respectively (see text and ref.[55,56]).

**Liver biopsy should be considered in patients with ferritin >1,000 µg/L or signs of liver damage.

***Exceptions may occur, e.g. in HC patients with diagnosis delayed after the appearance of liver cirrhosis, in whom anemia may be observed because of hypersplenism or gastrointestinal bleeding, or subjects with beta-thalassemia trait, whose coexistence is not rare in Mediterranean countries.

Table 2. Combined pathogenic allele frequency for HC genes in the 1000 Genomes Project (1000G), Exome Sequencing Project (ESP), and Exome Aggregation Consortium (ExAc) datasets (modified by Wallace et al.)[19]

Gene	1000G	ESP6500	ExAc	Geographical distribution
HFE (p.Cys282Tyr)	0.013	0.048	0.0324	Highest prevalence in Northern Europe
HFE (non-p.Cys282Tyr)	0.001	0.0002	0.000307	
HJV		0.00074	0.000316	Highest prevalence in Southern Asia
TFR2	0.0004	0.0003	0.000102	Most frequent among non-Finnish European populations
HAMP	0.0002		0.0000165	Several populations
SLC40A1	0.0008	0.0009	0.00034	Several populations (highest prevalence among Africans)

Table 3. Former classification of HC

Classification	Gene involved and location	Inheritance	TSAT	Other clinical features
Type 1	*HFE;* chr.6	AR	Increased	Adult-onset; more severe in males; highly variable clinical expression, with predominant liver damage and arthritis
Type 2A	*HJV* (hemojuvelin); chr.1	AR	Increased	Earlier onset (e.g.<30 years old); similar severity in both sexes; prevalent cardiac and endocrine involvement
Type 2B	*HAMP* (hepcidin); chr.19	AR	Increased	Earlier onset (e.g.<30 years old); similar severity in both sexes; prevalent cardiac and endocrine involvement
Type 3	*TFR2* (transferrin receptor 2); chr.7	AR	Increased	Very rare (look for parental consanguinity); clinically similar to Type 1, with an earlier onset
Type 4A	*SLC40A1* (ferroportin); chr.2	AD	Low-normal	Adult-onset; IO in the spleen; mild anemia; possible low tolerance to venesection
Type 4B	*SLC40A1* (ferroportin); chr.2	AD	Increased	Very rare; in general, clinically similar to Type 1, but more severe/early onset forms are reported

842 **Table 4. New classification of HC proposed by the working group**

843

Novel classification	Molecular pattern	Note
HFE-related	*p.Cys282Tyr* homozygosity or compound heterozygosity of *p.Cys282Tyr* with other rare *HFE* pathogenic variants[106-109] or *HFE* deletion[110]	Low penetrance; consider presence of host-related or environmental cofactors for IO In subjects with other HFE genotypes (e.g. *p.Cys282Tyr/His63Asp* compound heterozygosity or *p.His63Asp* homozygosity) consider second-line genetic testing for rarer variants.
Non HFE-related	Rare pathogenic variants in "*non-HFE*" genes: - *HJV*-related - *HAMP*-related - *TFR2*-related - *SLC40A1* (GOF)-related	Potentially, mutations in any hepcidin-regulatory gene may be causative (the effects of novel mutations should be confirmed through functional and epidemiological studies). Molecular subtypes characterization only at specialized centers, but the diagnosis of non-*HFE* related HC is sufficient to start phlebotomies at non-specialized centers*.
Digenic**	Double heterozygosity and/or double homozygosity/heterozygosity for mutations in two different genes involved in iron metabolism (*HFE* and/or *non-HFE*)	More commonly, *p.Cys282Tyr* mutation in *HFE* gene might coexist with mutation in other genes; rarely, both mutations involve *non-HFE* genes
Molecularly undefined	Molecular characterization (still) not available after sequencing of known genes (provisional diagnosis)	Patients should be referred (or DNA should be sent) to specialized centers

844 *Providing that iron overload is confirmed by MRI. If this is not accessible, close monitoring of Hb level is
845 needed to avoid the occurrence of anemia.
846 ** Caution is needed to interpret as digenic inheritance results from NGS outputs reporting several variants
847 in gene panels. Whenever possible, strict criteria for defining pathogenic variants should be adopted, and
848 corroborated by family segregation and/or functional studies.
849

Figure 1. Hepcidin regulation by iron. Increase in transferrin saturation induces hepcidin transcription via the BMP/SMAD signaling pathway. Diferric transferrin binds to TfR2, while BMP6 and BMP2 secreted by liver sinusoidal endothelial cells (LSECs) bind to BMP receptors on hepatocytes. These events trigger phosphorylation of regulatory SMAD1/5/8, recruitment of SMAD4, and translocation of the SMAD complex to the nucleus for activating hepcidin transcription upon binding to BMP/SMAD responsive element in the *HAMP* promoter. BMPs can be trapped by ERFE, leading to hepcidin inhibition in iron loading anemias. Efficient iron signaling requires the BMP co-receptor HJV and the protein HFE, and is negatively regulated by the transmembrane serine protease matriptase-2 (TMPRSS6). The complex molecular pathogenesis of HC reflects the numerous proteins involved in regulation of the hepcidin-ferroportin axis.

Figure 2. Proposal of an algorithm for the diagnosis of HC, from clinical/biochemical and imaging studies to molecular confirmation. Important note: in Caucasians, *HFE* genotyping is indicated with the specific purpose of detecting *p.Cys282YTyr* homozygosity and, if confirmed, to recommend appropriate preventive treatment by phlebotomies. Asian, African, and native American subjects with defined HC phenotype could be directly referred to second-level genetic test. In populations with a frequent component of northern European ancestry, such as African-Americans and Hispanics, there may be still a role for *HFE* genetic testing.

Figure 3. Iron homeostasis in normal conditions (A) and mechanisms leading to iron accumulation in HC (B) and in iron-loading anemias (C: non-transfusion dependent; D: transfusion-dependent). In HC, iron hyperabsorption through the portal vein leads to iron accumulation in liver parenchymal cells, initially with a typical portal-central gradient (see histology) and sparing of macrophages (Kupffer cells). In non-transfusion dependent anemias with ineffective erythropoiesis hepcidin insufficiency is also central to the pathogenesis of IO, but it is due to suppression by soluble factors (e.g. ERFE) produced by ineffective/expanded erythroblasts, rather than to a genetic defect in pathways regulating hepcidin synthesis. In transfusion dependent anemias, regular red blood cells (RBCs) transfusions represent the major contributing factor to IO; in these conditions, hepcidin is relatively upregulated by iron, but fluctuates in response to intermittent erythropoiesis suppression by transfusions.

Figure 1

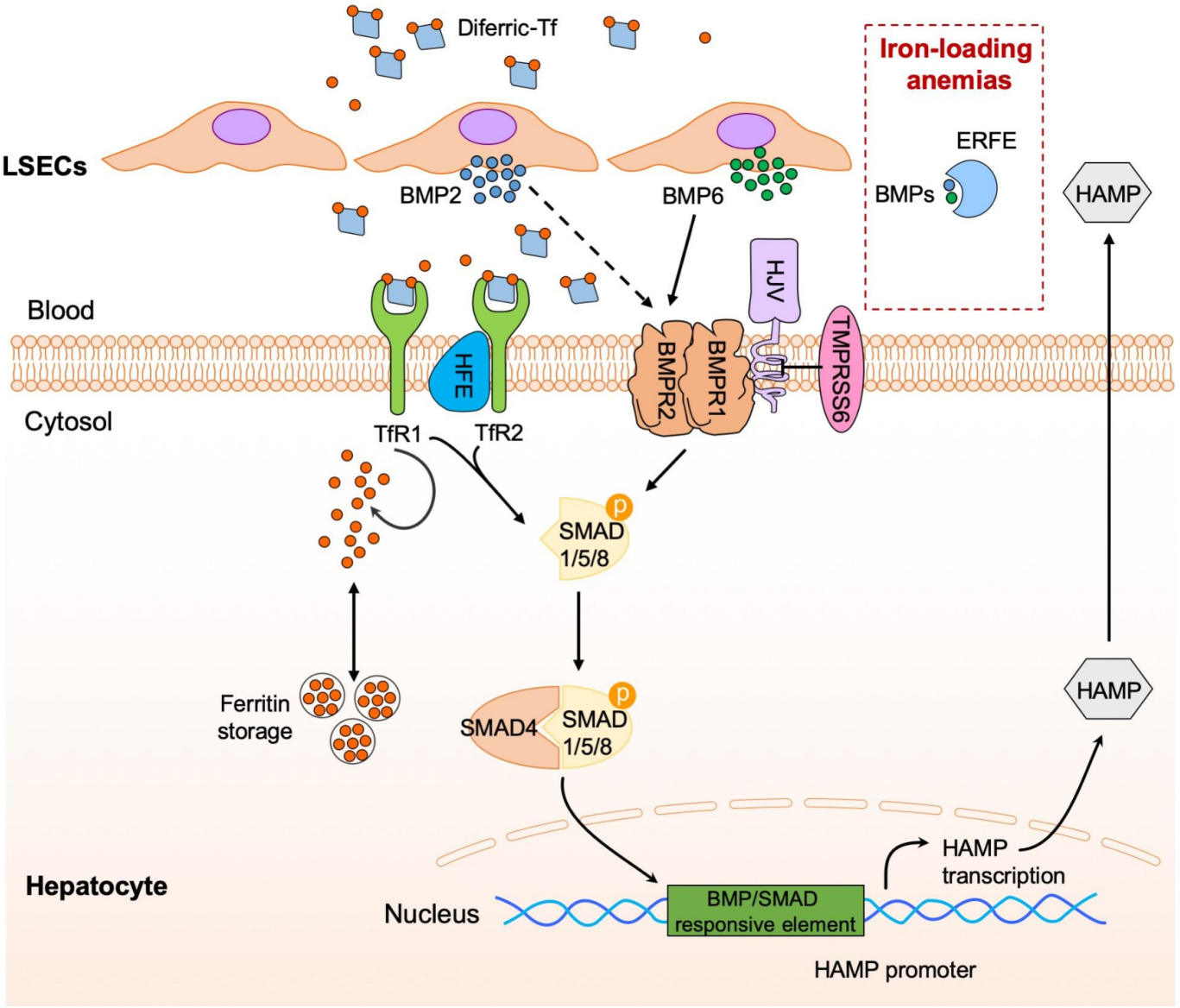

Figure 2

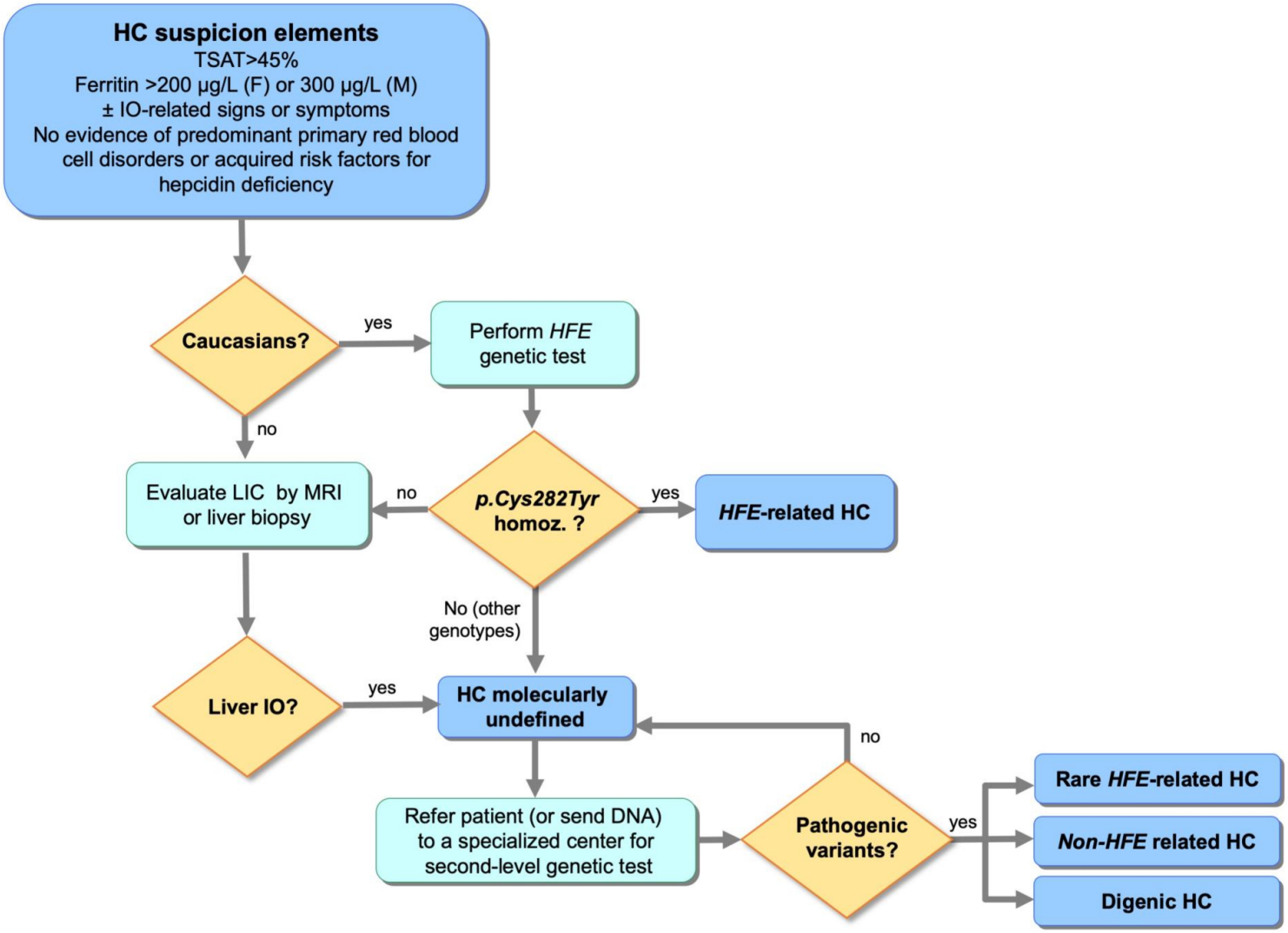

Figure 3